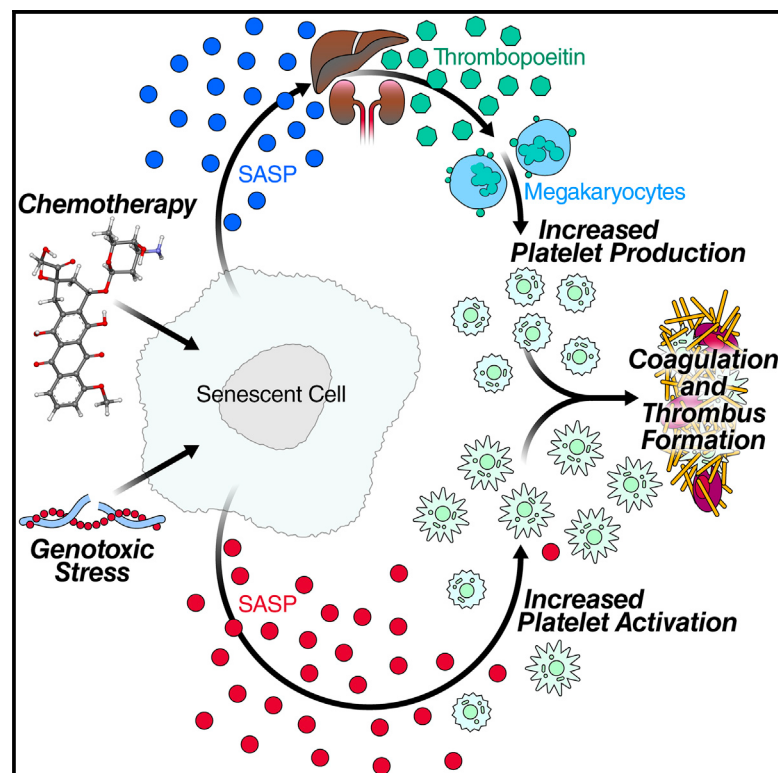


Cell Reports

SILAC Analysis Reveals Increased Secretion of Hemostasis-Related Factors by Senescent Cells

Graphical Abstract



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In Brief

Wiley et al. identify new proteins secreted by senescent cells using unbiased proteomics, including factors involved in hemostasis. Secretions of senescent cells increase human platelet activation, while eliminating senescent cells prevents increases in platelet counts and activation in mice treated with chemotherapy. Senescent cells may thus underlie clotting predisposition.

Highlights

- An unbiased SILAC screen identifies >300 proteins secreted by senescent human cells
- Forty-four of these proteins have reported roles in hemostasis
- Conditioned media from senescent cells promotes platelet activation
- Eliminating senescent cells ameliorates pro-coagulation side effects of doxorubicin



SILAC Analysis Reveals Increased Secretion of Hemostasis-Related Factors by Senescent Cells

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SUMMARY

Cellular senescence irreversibly arrests cell proliferation, accompanied by a multi-component senescence-associated secretory phenotype (SASP) that participates in several age-related diseases. Using stable isotope labeling with amino acids (SILACs) and cultured cells, we identify 343 SASP proteins that senescent human fibroblasts secrete at 2-fold or higher levels compared with quiescent cell counterparts. Bioinformatic analysis reveals that 44 of these proteins participate in hemostasis, a process not previously linked with cellular senescence. We validated the expression of some of these SASP factors in cultured cells and *in vivo*. Mice treated with the chemotherapeutic agent doxorubicin, which induces widespread cellular senescence *in vivo*, show increased blood clotting. Conversely, selective removal of senescent cells using transgenic p16-3MR mice showed that clearing senescent cells attenuates the increased clotting caused by doxorubicin. Our study provides an in-depth, unbiased analysis of the SASP and unveils a function for cellular senescence in hemostasis.

INTRODUCTION

Cellular senescence permanently arrests cell proliferation in response to a variety of stresses, including DNA damage, mitochondrial dysfunction, and oncogene activation (Campisi, 2013; Wiley et al., 2016). Senescent cells participate in a wide range of biological processes (Childs et al., 2015; Muñoz-Espín and Serrano, 2014), playing beneficial or deleterious roles, depending on the triggers of senescence, as well as the tissue and cell types. Senescent cells accumulate with age and drive a variety

of age-related pathologies, at least in mice (Baar et al., 2017; Baker et al., 2016; Chang et al., 2016). The senescence response can impair tissue regeneration (Chang et al., 2016; Jeon et al., 2017) and promote cancer cell proliferation and invasion (Krtolica et al., 2001; Laberge et al., 2015). In contrast, senescence suppresses tumorigenesis (Priour and Peeper, 2008), accelerates wound healing (Demaria et al., 2014), and fine-tunes embryogenesis (Muñoz-Espín et al., 2013; Storer et al., 2013). Although the mechanisms are incompletely understood, senescent cells exert some of these effects by secreting numerous growth factors, cytokines, chemokines, and proteases, known as the senescence-associated secretory phenotype (SASP) (Acosta et al., 2013; Coppé et al., 2008; Kuilman et al., 2008).

To better understand the biological consequences of cellular senescence, we embarked on a comprehensive, unbiased method to profile the SASP using stable isotope labeling with amino acids (SILACs), a mass spectrometry-based technique that uses non-radioactive, stable isotope labeling of cells. Surprisingly, this screen revealed several SASP factors, expressed by cells induced to senesce by DNA damage, that were predicted to participate in hemostasis.

Hemostasis is a tightly regulated, complex process that prevents excessive bleeding after injury. Hemostasis has primary and secondary major components. Primary hemostasis forms a platelet plug at the injured site. Upon vascular damage, platelets adhere to the injured vessel, become activated, and aggregate to arrest bleeding (Tanaka et al., 2009; Versteeg et al., 2013). However, because the primary plug is not stable, secondary hemostasis then stabilizes the clot by depositing insoluble fibrin, which is generated by the coagulation cascade. These two processes happen simultaneously and interact in a dynamic fashion. Multiple anti-clotting mechanisms, such as circulating anticoagulants and fibrinolysis, which removes the plug after the vessel is repaired, also play critical roles in hemostasis (Tanaka et al., 2009; Versteeg et al., 2013). An impaired balance between pro- and anti-elements of coagulation can lead to pathological bleeding or, conversely, thrombosis.



An increased risk for thrombosis is a common age-associated complication and major risk factor for mortality (Sepúlveda et al., 2015). Thrombosis is also a common side effect of radio- and genotoxic chemotherapies, which significantly increase the risk for post-therapy mortality and decrease quality of life (Bosch et al., 2014; Vergati et al., 2013). Though it is known that DNA-damaging cancer therapies induce cellular senescence (Demaria et al., 2017; Ewald et al., 2010), an association between cellular senescence and thrombosis has not yet been made.

Our finding that the secretion of a subset of hemostasis-related factors increases significantly when human fibroblasts undergo senescence suggests that senescent cells may contribute to the etiology of age- and genotoxic therapy-induced thrombosis. We show that senescent cells promote blood clotting in mice treated with the genotoxic anti-cancer chemotherapeutic doxorubicin, whereas removing senescent cells from such mice attenuates clotting. Furthermore, the pro-coagulation effect of senescent cells is associated with both increased platelet number and reactivity. Our findings identify an additional function of senescent cells and provide potentially important insights into genotoxic therapy-induced thrombosis.

RESULTS

SILAC Analysis Identifies a Potential Role for the SASP in Hemostasis

To obtain an unbiased profile of the SASP, we used SILACs to analyze conditioned media (CM) from quiescent and senescent primary human foreskin (HCA2) fibroblasts. Figure 1A and Figure S1A show the timeline for $^{13}\text{C}_6$ lysine and $^{13}\text{C}_6$ arginine labeling of quiescent cells and the schedule for inducing senescence in unlabeled cells by ionizing radiation (IR) and CM collection. We analyzed three independent biological replicates for each condition and confirmed the senescent and quiescent states by SA- β -gal staining and BrdU labeling (Figures S1B–S1C). The proteins we identified, and the magnitude of changes in secreted levels between quiescent and senescent cells, are shown in Tables S1A–S1C. Overall, we detected 1,047 proteins, identified by at least two peptides per protein, with 99% confidence (Table S1A).

Among the SASP factors we identified, 343 showed a significant (>2-fold) increase in abundance in CM from senescent, compared with quiescent, cells (Table S1B). Reactome bioinformatic analyses showed that, notably, 5 of the top 25 pathways (highlighted in Figure 1B) were associated with hemostasis, blood clot formation, or clot dissolution. The SASP factors predicted to be hemostasis related by Reactome, and the magnitude of their changes, are presented as two heatmaps in Figure 1C. The left heatmap shows factors that are the most highly elevated (most yellow), and the right shows the second most highly elevated. Additional bioinformatic analyses identified the molecular functions, cellular components, and biological processes in which they are known to participate (Figures S1D–S1F). Together, the results suggest the SASP might include hemostasis-related factors.

We also observed increases in extracellular matrix (ECM) proteins in our initial dataset, which might warrant additional study (Figure 1B; Figures S1D–S1F). However, the concurrent

increased secretion of proteinases may have artificially increased the abundance of peptides derived from the ECM. For this reason, we chose to focus on hemostasis.

Validation of SILACs Results in Cultured Cells

To validate the SILAC results, we collected CM from three independent cultures of quiescent or IR-induced senescent HCA2 cells and determined by western blotting the levels of 11 hemostasis-related factors identified by SILACs (Figure 1D). These factors included vinculin (VCL), tissue inhibitor of metalloproteinase 1 (TIMP1), tissue factor pathway inhibitor (TFPI), thrombospondin 1 (THBS1), calumenin (CALU), serpin E1 (SERPINE1, PAI1), serpin E2 (SERPINE2), serpin B2 (SERPINB2, PAI2), serpin B6 (SERPINB6), filamin A-alpha (FLNA), and EH domain containing 2 (EHD2). All were present at increased levels in CM from senescent, compared with quiescent, cells in all three replicates, consistent with the SILAC results. These factors play important roles in regulating hemostasis. For example, THBS1 is a major protein released from platelets during activation, where it stimulates platelet aggregation by blocking nitric oxide/cGMP signaling (Isenberg et al., 2008).

Because thrombosis is a common unwanted side effect of genotoxic cancer treatments (Ashrani et al., 2016; Huang et al., 2011; Zagar et al., 2016), we compared factors from quiescent cells and cells induced to senesce by doxorubicin (DOXO), a DNA-damaging chemotherapeutic. Elevated secretion of many SASP factors correlates with increased mRNA abundance of the respective genes (Coppé et al., 2008). Indeed, mRNAs encoding several hemostasis-related factors increased after DOXO treatment (Figure 2A). We confirmed the induction of hemostasis-related genes upon senescence in another primary human fibroblast strain, WI-38 (Figure S2A). Because blood is more likely to be exposed to senescent vascular cells, we also tested senescent human umbilical vein endothelial cells (HUVECs) for ability to produce these factors (Figure S2B). In agreement with previous work (Coppé et al., 2008), the patterns were not identical. However, the expression of several major hemostasis factors increased upon irradiation of HUVECs, similar to fibroblasts, though *PLAU*, *PLAUR*, and *FLNA* were each lower in senescent HUVECs. Moreover, western blots of CM from DOXO-induced senescent HCA2 cells showed increased hemostatic SASP factors relative to quiescent controls, similar to IR-induced senescence (Figure 2B). Finally, DOXO can also deplete mtDNA. We therefore tested the levels of these factors in mitochondrial dysfunction-associated senescence (MiDAS) induced by mtDNA depletion (Wiley et al., 2016). Many hemostasis-related factors were also elevated in MiDAS (Figure 2C), indicating that multiple drivers of senescence potentially affect hemostasis.

Senescence-Stimulated Induction of Hemostasis-Related Genes *In Vivo*

To determine whether the senescence-associated increased expression of hemostasis-related genes occurs *in vivo*, we used p16-3MR transgenic mice, which allow the detection of senescent cells by luminescence and their removal by ganciclovir (GCV) (Demaria et al., 2014). Following systemic administration of DOXO, senescent cells increased, as measured by elevated luminescence and $p16^{\text{INK4a}}$ mRNA levels (Figures S3A and

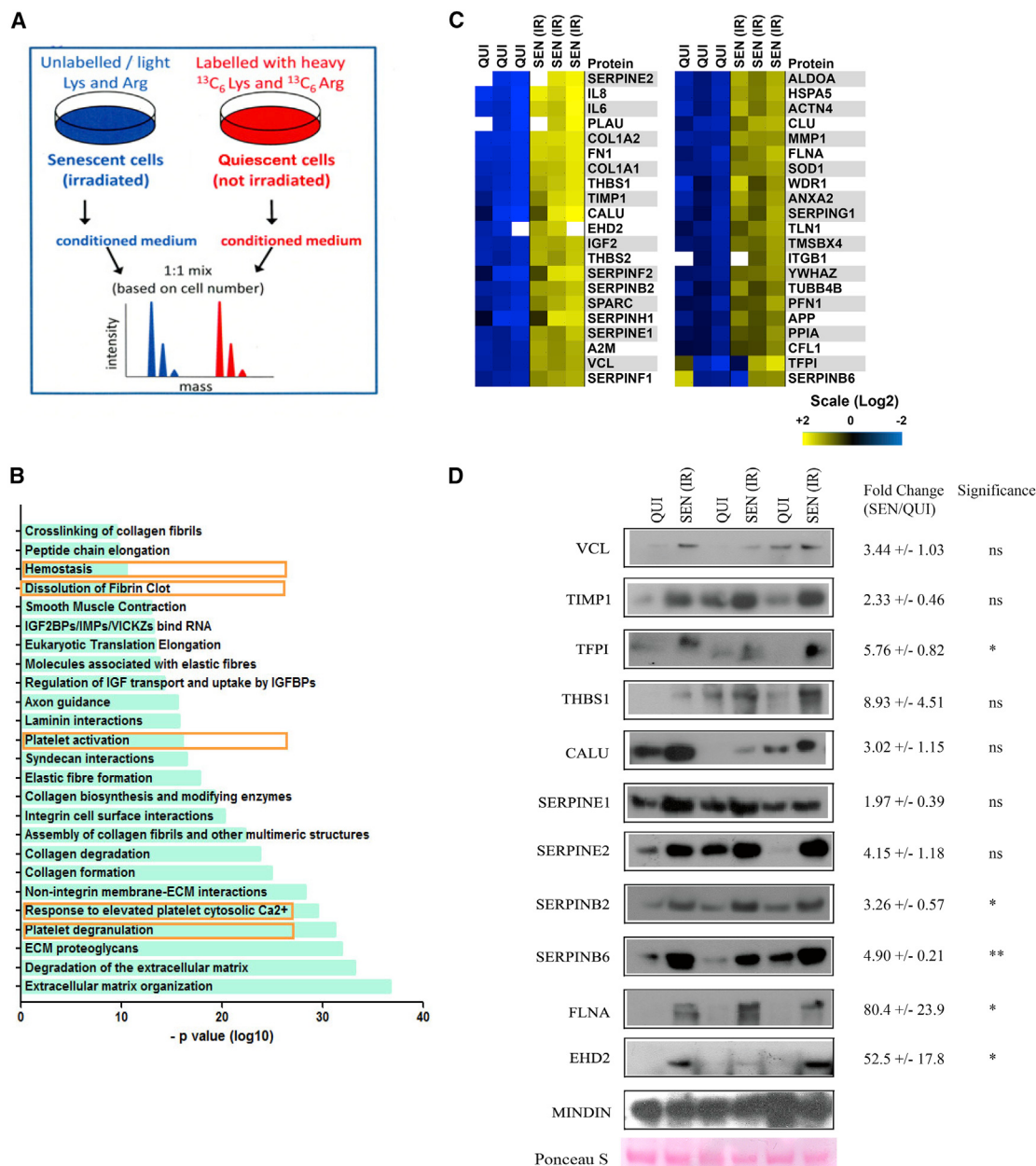


Figure 1. SILAC Analysis Suggests a Role for the SASP in Hemostasis

(A) A schematic outlining the strategy for SILAC-based identification of SASP proteins. See [Figure S1](#) for time lines of how cells were made quiescent or senescent and how quiescent and senescent conditioned media (CM) were produced.

(B) Reactome pathway analysis of proteins secreted at >2-fold levels by senescent compared with quiescent HCA2 cells. Pathways related to hemostasis are identified by boxes.

(C) Heatmap showing individual levels of proteins, identified by SILACs, that participate in hemostasis and are secreted by quiescent (QUI) and senescent (SEN [IR]) HCA2 normal human fibroblasts.

(D) CM were collected from quiescent (QUI) and senescent (SEN [IR]) cells, concentrated, and analyzed using western blotting for the indicated SASP factors. Cells remaining on the plate at the time of CM collection were counted for normalization. Mindin protein levels and Ponceau S staining were used as loading controls.

Fold changes are mean $\pm$ SEM. * $p < 0.05$ and ** $p < 0.01$.

See also [Figure S1](#) and [Table S1](#).

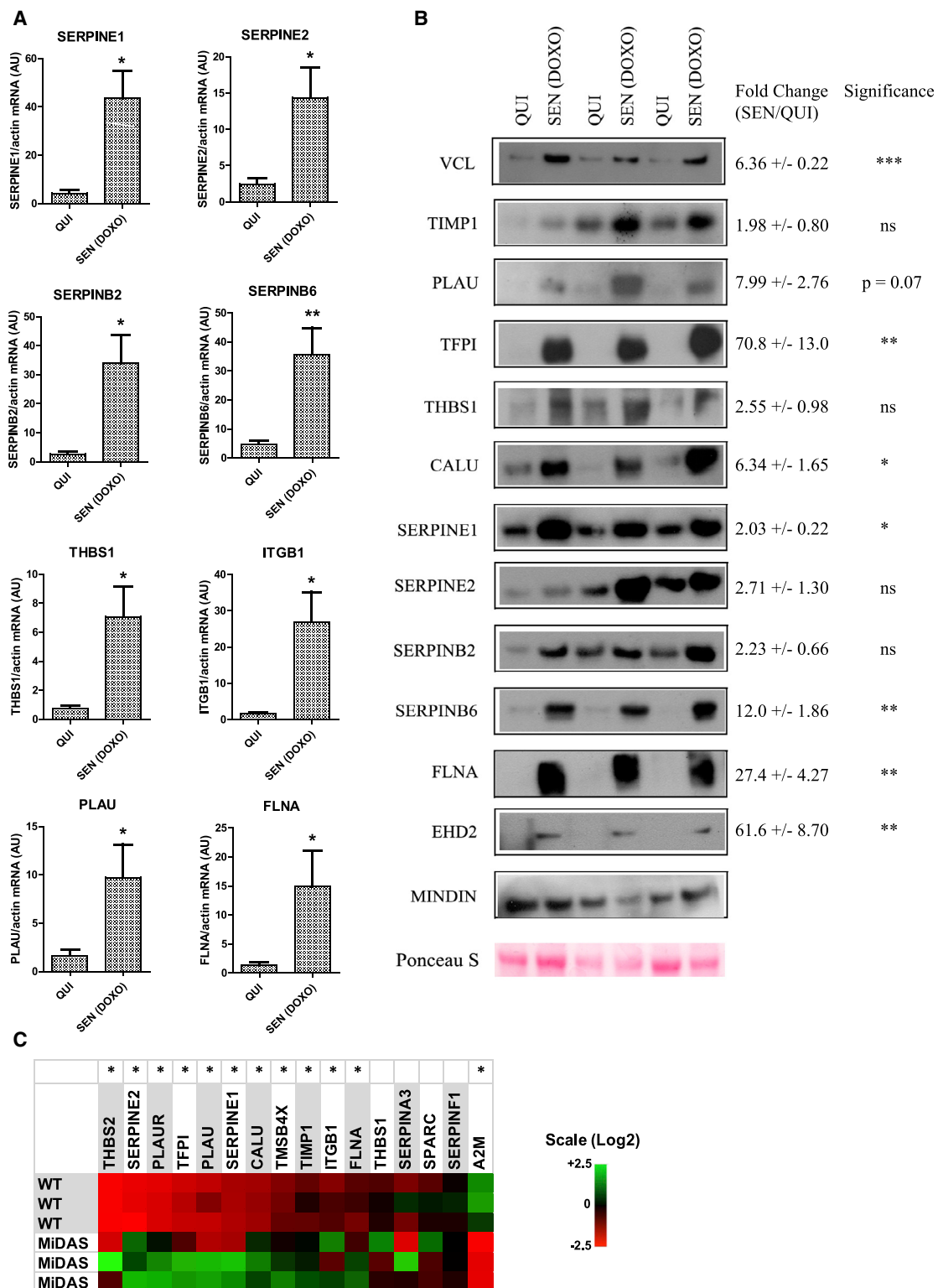


Figure 2. Doxorubicin Induces Expression of Hemostasis Factors

(A) mRNA was extracted from HCA2 cells made quiescent (QUI) or induced to senescence by doxorubicin (SEN [DOXO]), and analyzed using qPCR for the indicated genes, using actin mRNA for normalization. The data are shown in a.u., and the bars show mean $\pm$ SEM. *p < 0.05 and **p < 0.01, two-tailed, unpaired t test.

(legend continued on next page)

S3B) in the liver, a key organ that generates hemostasis-related factors, and these senescent cells were efficiently eliminated by GCV (Demaria et al., 2017). Importantly, mRNA levels of the hemostasis-related genes *Serpine1*, *Serpine2*, *Thbs1*, *Plau*, *Flna*, and *Timp1* were also elevated by DOXO and reduced by GCV (Figure 3A).

We extracted platelet-poor plasma from each treatment group and measured *Thbs1* (Figure 3B, left panel), *Serpine1* (Figure 3B, right panel), uPA (Figure S3C, right panel), and *Timp1* (Figure S3C, left panel). Each factor increased after DOXO treatment and declined upon removal of senescent cells by GCV. Moreover, mRNAs encoding senescence (*p16^{INK4a}* and *Il6*) (Figure S3D) and hemostasis-related factors were elevated in livers of aged mice (24–25 months old compared with 4–6 months old) (Figures S3D and S3E).

Senescent Cells Lower Tail Bleed Times

To determine whether senescent cells affect hemostasis *in vivo*, we analyzed tail bleeding times in DOXO- or PBS-treated p16-3MR mice receiving GCV or vehicle. DOXO-treated mice showed significantly shorter bleeding times relative to control mice. Importantly, after eliminating senescent cells, tail bleeding times were similar to those of control mice (Figure 4A). We also determined the amount of blood lost in the tail bleeding time assay by measuring hemoglobin levels (Figure 4B). Consistent with the tail bleeding time assays, DOXO-treated mice had the lowest blood loss, which was prevented by GCV. Because red blood cell counts were similar among the treatment groups (Table S2), lower hemoglobin in DOXO-treated mice was not due to anemia but rather to faster clotting.

Finally, platelet counts increased in response to DOXO and returned to baseline levels when DOXO-treated mice were treated with GCV, indicating that the increased platelet counts were senescence dependent (Figure 4C). Plasma from DOXO-treated mice showed increased thrombopoietin (Tpo), which drives thrombopoiesis, and this increase was also senescence dependent (Figure 4D). Despite this observation, we found no evidence for changes in the burden of senescent cells or hemostatic factors in bone marrow derived from treated mice (Figure S4A), suggesting that the effect of senescent cells on coagulation is not driven by changes in the marrow environment. Consistent with this suggestion, we found no changes in maturation when megakaryocytes were removed from the marrow of each group and cultured separately, indicating that secreted factors, rather than megakaryocytes themselves, drove this effect (Figure S4B).

Senescent Cells Sensitize Platelets to Activation

Clotting is regulated primarily by platelet activation and the coagulation cascade (Semple et al., 2011). To determine whether senescent cells affect platelet activation *in vivo*, we compared platelets in the blood of DOXO- or vehicle-treated

p16-3MR mice that were subsequently given GCV or vehicle. We used collagen to activate the platelets and measured P-selectin positivity (Figure 4E) and exposed phosphatidylserine (assessed by annexin V binding) (Figure 4F), markers of platelet activation, using flow cytometry. Collagen activated a greater number of platelets from DOXO-treated mice relative to controls; importantly, GCV blunted this increase in activated platelets.

To determine whether the SASP affects platelet activation, we incubated freshly isolated human platelets with CM from the quiescent or IR-induced senescent human cells (described in Figure 1) and the platelet activator collagen. We then measured platelet-released ATP, a marker of activation (Beigi et al., 1999). Platelets exposed to CM from senescent cells showed greater activation than platelets exposed to quiescent cell CM (Figure 4G), indicating that the SASP can prime platelets for activation.

Finally, we asked whether senescent cells affect the coagulation cascade. We measured activated partial thromboplastin time (APTT) and prothrombin time (PT) in each treatment group (Quality Veterinary Laboratory, Davis, CA). There were no significant differences between DOXO- and PBS-treated p16-3MR mice (Figures S4C and S4D), suggesting that senescent cells do not affect the coagulation cascade. Because many factors identified in our analysis can accelerate the coagulation cascade (e.g., serpins, uPA), it is likely that the factors that promote platelet activation (e.g., THBS1) play a more important role in the promotion of hemostasis by senescent cells. Together, the data indicate that senescent cells influence clotting time primarily by increasing platelet number and sensitizing them to activation.

DISCUSSION

The SASP can promote cancer cell proliferation and metastasis (Coppé et al., 2008; Krtolica et al., 2001; Laberge et al., 2015), reinforce senescence (Acosta et al., 2013), and optimize wound healing (Demaria et al., 2014) and embryonic development (Muñoz-Espín et al., 2013; Storer et al., 2013). Thus, investigating the complete SASP profile is critical to determine the biological functions of senescent cells. A previous study identified SASP factors from oncogene-induced senescent cells using SILACs (Acosta et al., 2013). We found 32 proteins in common between this study and ours, accounting for ~10% of the total SASP factors we identified and ~25% of the secretome identified by the previous study. The differences between these SASP profiles may be due to the differences in cells, collection time, sample handling protocol, and senescence stimuli. Recent data suggest that cellular senescence is not a singular homogeneous state but rather an evolving heterogeneous phenotype regulated with spatiotemporal specificity (Demaria et al., 2014; Hoare et al., 2016). Thus, future genomic and proteomic studies will be needed to explore different stimuli, time points, and cell types.

(B) CM were collected from cells treated as in (A), concentrated and analyzed using western blotting for the indicated SASP factors. Cells were counted for normalization. Mindin protein levels and Ponceau S staining were used as loading controls.

(C) IMR-90 fibroblasts were depleted of mtDNA by culture in ethidium bromide until senescent (21 days) (labeled as MiDAS). RNA was extracted and analyzed using qPCR for the indicated factors. Heatmap represents individual values for three experiments.

**p* < 0.05, one-way ANOVA.

See also Figure S2.

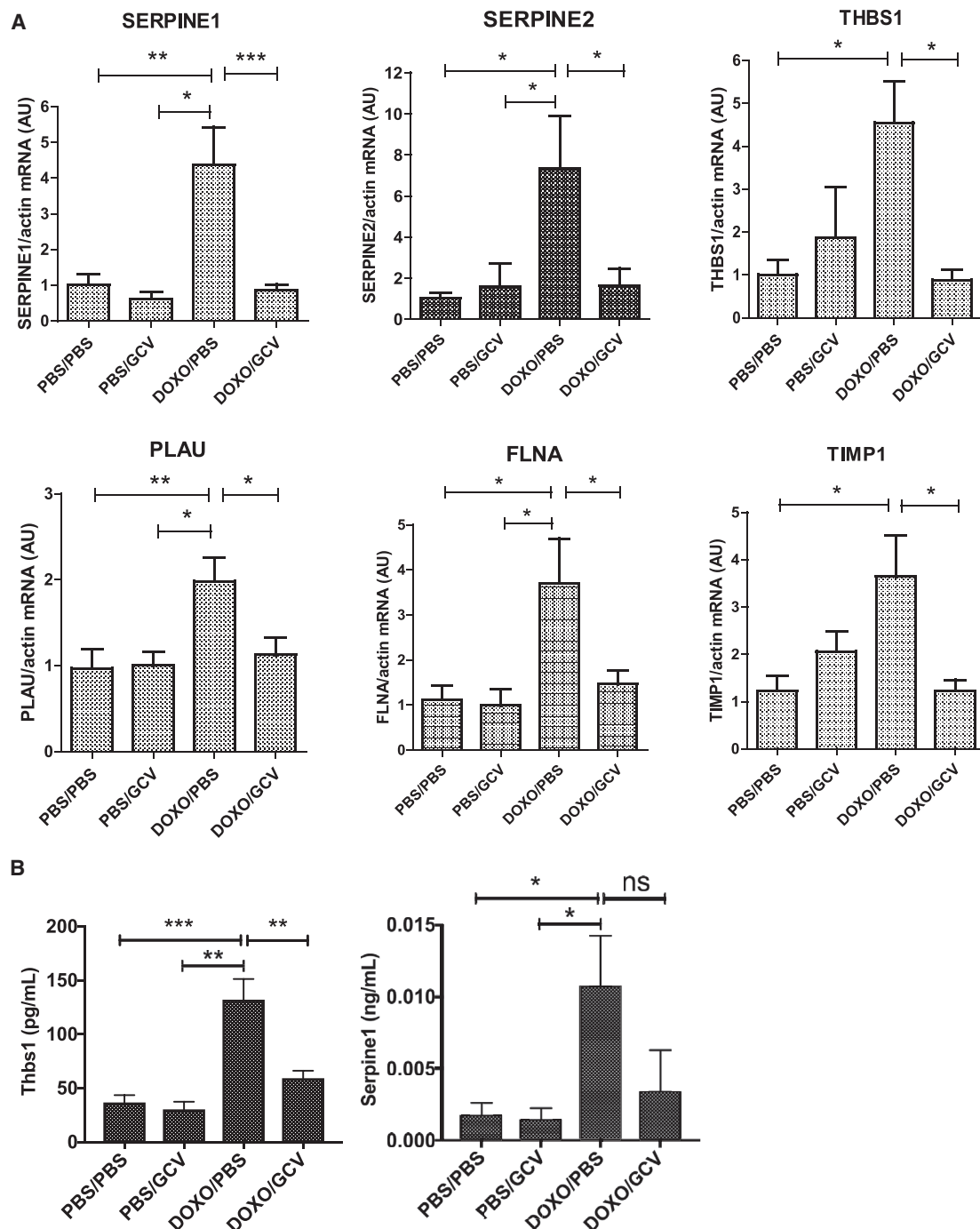


Figure 3. In Vivo Detection of the Hemostatic SASP

Young (4–7 months old) p16-3MR mice were given PBS (control) or DOXO (10 mg/kg) by intraperitoneal (i.p.) injection. After 7 days, they were also given PBS (control) or ganciclovir (GCV; 25 mg/kg, i.p.) once per day for 5 consecutive days to clear senescent cells, as described (Demaria et al., 2014).

(A) RNA was extracted from the liver of treated mice and analyzed using qPCR for mRNA levels of the indicated genes. a.u. values were normalized to actin mRNA.

(B) Platelet-poor plasma from each treatment group was analyzed using ELISA for Thbs1 or Serpine1.

All bars show mean $\pm$ SEM. * $p < 0.05$, ** $p < 0.01$, and *** $p < 0.001$, one-way ANOVA with Tukey's post test. $n \geq 7$ for all experiments.

See also Figure S3.

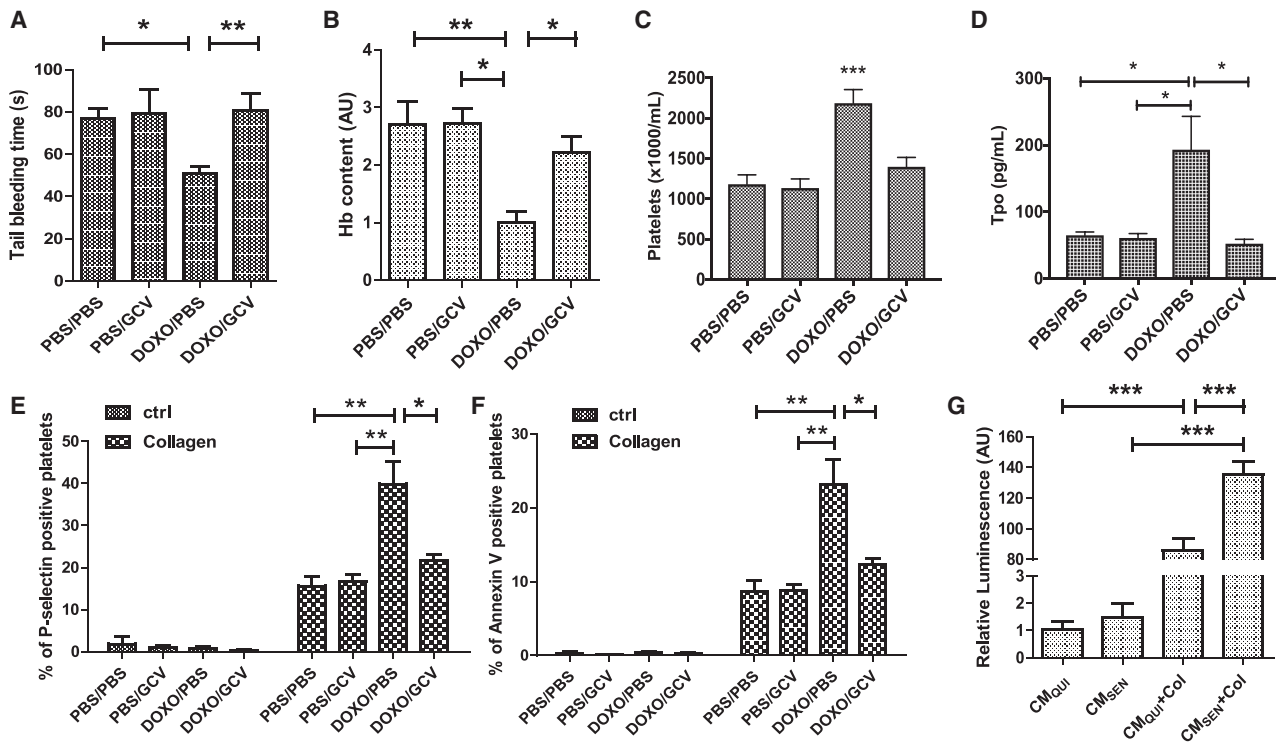


Figure 4. Senescent Cells Reduce Tail Bleeding Time and Prime Platelet Activation

(A–F) Young (4–7 months old) p16-3MR mice were given PBS or DOXO followed by PBS or GCV as described for Figure 3.

(A) Transected tails were submerged in 37°C saline, and time to bleeding cessation was measured. The data are expressed as mean $\pm$ SEM for 15 to 30 mice per group.

(B) Blood loss was quantified by measuring the hemoglobin (Hb) content of saline from the tail bleeding assays in a.u.

(C) Platelet counts from whole blood extracted from mice.

(D) Thrombopoietin ELISA from plasma obtained from the indicated mice.

(E and F) Whole blood was isolated from the animals, washed, stimulated with collagen, and analyzed using flow cytometry for cell surface P-selectin (E) and exposed phosphatidylserine (F). Data are expressed as the percentage of P-selectin-positive cells (E) and annexin V (which specifically binds phosphatidylserine) positive cells (F).

(G) Human platelet-rich plasma was incubated with CM from quiescent (CM_{QUI}) or IR-induced senescent (CM_{SEN}) cells without or with collagen (+Col) for 10 min. Released ATP was measured by luminescence over 5 min. The value for CM_{QUI} was set arbitrarily at 1.

Bar graphs show mean $\pm$ SEM. *p < 0.05, **p < 0.01, and ***p < 0.001 by one-way ANOVA with Tukey's post test for all experiments.

See also Figure S4 and Table S2.

Our SILAC results suggest a role for the SASP in hemostasis. Indeed, senescent cells promoted blood clotting, most likely by sensitizing platelets to agonists rather than activation of the coagulation cascade. Platelet activation is associated with thrombotic disease (Gurbel et al., 2016; Ruggeri, 2002), and anti-platelet therapy has been used to prevent and treat cerebrovascular, coronary, and peripheral arterial diseases (Michelson, 2010; Steinhubl et al., 2014; Tran and Anand, 2004). Chemotherapy is a major risk factor for thrombotic disorders, including stroke (Huang et al., 2011), cardiovascular disease (Zagar et al., 2016), and venous thromboembolism (Ashrani et al., 2016). Although there is a 4.1-fold greater risk for thrombosis in cancer patients, chemotherapy increases this risk to 6.5-fold (Heit et al., 2000), with as yet no clear explanation. DOXO was previously shown to increase platelet activation *in vitro*, but the role of senescent cells in this process was not studied (Kim et al., 2011). Because genotoxic chemotherapies are known to induce cellular senescence (Demaria et al., 2017;

Ewald et al., 2010), our findings suggest that chemotherapy-induced senescence could potentiate blood clotting, possibly by modifying platelet function. They also suggest that SASP-suppressing adjuvant therapies might reduce this side effect of chemotherapy.

The SASP is a multi-component phenotype that entails the secretion of dozens of biologically active factors (Acosta et al., 2013; Coppé et al., 2008; Kuilman et al., 2008). Our findings suggest that the SASP also regulates homeostasis-related factors, among other biological activities. However, the precise mechanisms by which senescent cells sensitize platelets for activation remain to be determined. Because senescent cells accumulate with age, it is tempting to speculate that the SASP is at least partly responsible for the thrombotic events that increase with age. These indications are potential targets for the development of senolytic drugs, which can selectively eliminate senescent cells (Chang et al., 2016), similar to the action of GCV in p16-3MR mice.

STAR★METHODS

Detailed methods are provided in the online version of this paper and include the following:

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SUPPLEMENTAL INFORMATION

Supplemental Information can be found online at <https://doi.org/10.1016/j.celrep.2019.08.049>.

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AUTHOR CONTRIBUTIONS

S.L., A.M.Z., and S.R.D. designed and conducted SILAC analyses with guidance from B.S., J.C., P.K., and B.W.G. C.D.W. and S.L. identified, designed, and conducted hemostasis experiments. R.A., N.B., T.R.O., and S.D.M. provided bioinformatic support and ontological analyses. S.L., C.D.W., C.L., M.D., J.-A.L.-D., and F.A. conducted animal experiments. C.K. analyzed bone marrow. H.G.K. performed platelet counts. C.D.W., S.L., J.-A.L.-D., and J.B. performed RNA analyses. S.L., C.D.W., P.-Y.D., B.S., J.C., and P.K. wrote and edited the paper.

DECLARATION OF INTERESTS

J.C. is a founder and shareholder of Unity Biotechnology, which aims to develop senolytic drugs. The other authors declare no competing interests.

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STAR★METHODS

KEY RESOURCES TABLE

REAGENT or RESOURCE	SOURCE	IDENTIFIER
Antibodies		
p-selectin	BD-PharMingen	553744
CALU	Abgent	AP12308b-ev
EHD2	Abgent	aw5422-EV
PLAU	Abgent	AP8161b-ev
SERPINE2	Abgent	AP12607c-ev20
FLNA	Bethyl	A301-134A-T
CD41	Biologend	133905
Annex-V-APC	Biologend	640920
MINDIN	R&D Systems	AF2609
SERPINB2	R&D Systems	MAB85501
SERPINB6	R&D Systems	AF4336
SERPINE1	R&D Systems	MAB1786
TFPI	R&D Systems	AF2974
THBS1	R&D Systems	AF3074
TIMP1	R&D Systems	AF970
VCL	R&D Systems	MAB6896
Chemicals, Peptides, and Recombinant Proteins		
Acetonitrile	Burdick & Jackson	015
Formic acid	Sigma-Aldrich	F0507
HPLC grade water	Burdick & Jackson	365
Dulbecco's modified eagle medium (DMEM)	Fisher Scientific	21-063-029
F-12K medium	ATCC	30-2004
Fetal bovine serum (FBS)	Fisher Scientific	16140
Endothelial cell growth supplement	BD Biosciences	#354006
Ethidium bromide	Thermo Scientific	15585-011
Doxorubicin	Sigma-Aldrich	D1515
Ganciclovir	Sigma-Aldrich	G2536
Penicillin/Streptomycin	Genesee Scientific	25-512
Iodoacetamide	Sigma-Aldrich	I1149
Dithiothreitol (DTT)	Sigma-Aldrich	43815
Ammonium bicarbonate	Sigma-Aldrich	09830
Urea	Sigma-Aldrich	GE17-1319-01
Sequencing Grade Modified Trypsin	Promega	V5111
L-Arginine-HCl, 13C6, 15N4 for SILAC	Thermo Scientific	89990
L-Lysine-2HCl, 13C6 for SILAC	Thermo Scientific	89988
Amicon® Ultra-15 Centrifugal Filter Unit	EMD Millipore	UFC900308
UltraPure 1M Tris-HCl, pH 8.0	Thermo Scientific	15568025
TSKgel® Amide-80 HPLC Column	Sigma-Aldrich	813071
Acclaim PepMap 100 C18 LC Columns	Thermo Scientific	160321
Collagen	Chrono-Log	385
CountBright Absolute Counting Beads	Thermo Scientific	C36950
Propidium iodide	Life Technologies	P3566
L-Glutamine	Life Technologies	25030081

(Continued on next page)

Continued

REAGENT or RESOURCE	SOURCE	IDENTIFIER
Recombinant Mouse Thrombopoietin Protein	R&D Systems	488-TO
Ponceau S solution	Sigma Life Science	P7170
Critical Commercial Assays		
Senescence Detection Kit	BioVision	K320
5-Bromo-2'-deoxy-uridine Labeling and Detection Kit	Roche	#11296736001
CHRONO-LUME®	Chrono-Log	395
Mouse THBS1 / Thrombospondin-1 ELISA Kit (Sandwich ELISA)	LifeSpan Biosciences	LS-F12876
Mouse PAI1 ELISA Kit	Abcam	ab197752
Mouse uPA ELISA Kit	Innovative Research	IMUPAKT
Mouse TIMP-1 Quantikine ELISA Kit	R&D Systems	MTM100
Mouse Thrombopoietin Quantikine ELISA Kit	R&D Systems	MTP00
ISOLATE II RNA mini kit	Bioline	BIO-52073
Direct-zol RNA MiniPrep Kit	Zymo	11-331
High Capacity cDNA Reverse Transcription Kit	Applied Biosystems	4368813
Deposited Data		
Raw data from this study (MSV000079131)	MassIVE	ftp://MSV000079131@massive.ucsd.edu (password:winter)
Experimental Models: Cell Lines		
HCA2 foreskin fibroblasts	Olivia Pereira-Smith	N/A
Wi38 fibroblasts	ATCC	CCL-75
HUVEC-C endothelial cells	ATCC	CRL-1730
IMR-90 fibroblasts	ATCC	CCL-186
Experimental Models: Organisms/Strains		
p16-3MR mouse in C57BL/6 background	Judith Campisi	N/A
Oligonucleotides		
RT-PCR Primers and UPL Probes	See Table S3	N/A
Software and Algorithms		
ProteinPilot	SCIEX Beta 4.5	Revision 1656
SwissProt	UniProt Knowledgebase	2014_01 release
The Database for Annotation, Visualization and Integrated Discovery	https://david.ncifcrf.gov/	v6.7
GO enrichment analysis	http://geneontology.org	2014 release
ImageJ	https://imagej.nih.gov/	1.52d 11 June 2018

LEAD CONTACT AND MATERIALS AVAILABILITY

Further information and requests for resources and reagents should be directed to and will be fulfilled by Pankaj Kapahi (pkapahi@buckinstitute.org). p16-3MR mice are shared for research and educational purposes under an MTA that will be discussed in good faith with the recipient.

EXPERIMENTAL MODEL AND SUBJECT DETAILS

Cell Culture

HCA2 are normal primary human neonatal foreskin fibroblasts (originally obtained from Dr. Pereira-Smith, University of Texas Health Science Center, San Antonio, TX). WI-38 and IMR-90 are normal primary human embryonic lung fibroblasts (obtained from the American Type Culture Collection). Fibroblasts were cultured in DMEM plus 10% FBS and penicillin/streptomycin, at 3% oxygen and 10% CO₂. HUVECs were obtained from the American Type Culture Collection and grown in F-12K medium plus 10% FBS, penicillin/streptomycin, heparin, and endothelial cell growth supplement (BD Biosciences #354006). Senescence was induced by exposing sub-confluent cultures to 10 Gy X-irradiation (IR) or 250 nM doxorubicin (DOXO); control cultures were mock irradiated or treated with vehicle. Quiescence was induced by culture in 0.2% serum for 3 days. Quiescent cells were used 3 d after mock irradiation or vehicle treatment; senescent cells were used 7-10 d after irradiation or DOXO treatment. Media were changed every other

day. On the 9th day, cells were incubated in serum-free medium and CM was collected 24 h later. For mitochondrial dysfunction-associated senescence (MiDAS), IMR-90 fibroblasts were cultured in the presence of ethidium bromide (100 ng/mL) until senescent, as described (Wiley et al., 2016).

Animal Experiments

All protocols were approved by the Buck Institution Animal Care and Use Committee. Young (4–7 months old) p16-3MR male mice were given PBS (vehicle) or DOXO (10 mg/kg) by intraperitoneal (i.p.) injection. Seven days later, they were given, also by i.p. injection, PBS (vehicle) or ganciclovir (GCV; 25 mg/kg) once per day for 5 consecutive days to clear senescent cells, as described (Demaria et al., 2014).

For tail bleeding assays, male mice (p16-3MR in C57BL/6, 3–5 months old) were anesthetized, and the tail was transected 5 mm from the tip with a razor blade. The bleeding tail was immersed in a 15 mL test tube containing saline pre-warmed to 37°C, and time to cessation of blood flow was measured. Bleeding times were determined when the bleeding stopped for 10 s. Bleeding times exceeding 6 min were noted as 6 min. The amount of bleeding was quantitated by measuring the hemoglobin content of blood collected into the saline using a Hemoglobin Assay Kit (Sigma Aldrich, St. Louis, MO), following the manufacturer's instructions.

METHOD DETAILS

Antibodies

Antibodies against the following proteins were used (see also [Key Resource Table](#)): **BD Pharmigen** – P-selectin (553744); **Abgent** – CALU (AP12308b-ev), EHD2 (aw5422-EV), PLAU (AP8161b-ev), SERPINE2 (AP12607c-ev20); **Bethyl** – FLNA (A301-134A-T); **Bio-legend** – CD41 (133905), Annex-V-APC (640920); **R&D Systems** – MINDIN (AF2609), SERPINB2 (MAB85501), SERPINB6 (AF4336), SERPINE1 (MAB1786), TFPI (AF2974), THBS1 (AF3074), TIMP1 (AF970), VCL (MAB6896).

SA- β -Gal and BrdU Labeling

We detected SA- β -Gal as described (Dimri et al., 1995), using a commercial kit (BioVision). Briefly, 30,000 cells per well were seeded on a 12-well plate. Cells were fixed using 500 μ L Fixing Buffer for 10 min, followed by three washes in PBS, and overnight incubation at 37°C in Staining Solution. Cells were visualized and counted using an inverted microscope.

DNA synthesis was determined using a modified BrdU labeling kit (Roche #11296736001). Cells were cultured in BrdU (20 μ M in DMSO) for 24 hours, fixed in 4% neutral-buffered formalin for 10 min, and washed 3 times in ice cold PBS. Permeabilization was performed for 30 min by incubation in 0.5% Triton X-100 in PBS. Nuclear DNA was digested by a combination of DNase I and exonuclease III for 1 hour, and BrdU-containing fragments were visualized by immunofluorescence.

Reagents and Standards

HPLC solvents, including acetonitrile and water, were obtained from Burdick & Jackson (Muskegon, MI). Reagents for protein chemistry, including iodoacetamide, dithiothreitol (DTT), ammonium bicarbonate, formic acid, and urea, were purchased from Sigma Aldrich (St. Louis, MO). Proteomics grade trypsin was from Promega (Madison WI). HLB Oasis SPE cartridges were purchased from Waters (Milford, MA). SILAC heavy lysine Lys-6 (13C6, 99%) and Arg-6 (13C6, 99%) were obtained from Thermo Scientific (Waltham, MA).

SILAC Sample Preparations

Non-senescent cells were grown in heavy SILAC medium, and senescent (irradiated) cells were incubated in light SILAC medium, both with 3 biological replicates (for details see [Figure S1A](#)). Quiescent HCA2 cells were cultured in DMEM medium supplied with L-Lys-¹³C₆ and L-Arg-¹³C₆ (heavy medium) for 6 passages to achieve complete label incorporation (~99%) before seeding (Day 0), according to the vendor's recommendation. At least 5 passages prior to the experiment, incorporation efficiency of the heavy labeled amino acids into proteins was assessed in a pilot experiment, where a small aliquot of intracellular protein was lysed, reduced, alkylated, and trypsin digested and subjected to MS analysis as described below. Heavy label incorporation into proteins from non-senescent cells was assessed as >99%. Senescent (irradiated) HCA2 cells were cultured in DMEM complete medium (light medium) (for details see [Figure S1A](#)).

For the proteomics, 24 h before harvesting, plated cells were washed 3 times with fresh fetal calf serum (FCS)-free medium for 10 min at 37°C each. The cells were then incubated in FCS-free medium (12 mL per flask) for 24 h at 37°C. At the end of the culture period, cells showed no evidence of apoptosis. CM was collected as described above, and centrifuged at 1,200 g for 5 min. The supernatant was transferred to fresh tubes. CM from quiescent and senescent cells were mixed based on equal cell numbers, and concentrated (~1:100 fold) using Millipore centrifugal filter units (MWCO 3.5 kDa). Samples were prepared for mass spectrometric analysis including removal of phenol red dye by buffer exchange with 50 mM TrisCl (pH8.0) (5 washing steps using spin columns at x 5000 g). CM were denatured with 6 M urea, reduced with 20 mM DTT (30 min at 37°C), alkylated with 50 mM iodoacetamide (30 min at RT), and digested overnight at 37°C with 1:50 enzyme:substrate ratio (wt/wt) of sequencing grade trypsin (Promega, Madison, WI) as described (Keshishian et al., 2007). Following digestion, samples were acidified with formic acid and desalted using HLB Oasis SPE

cartridges (Waters, Milford, MA). Protein/peptide recovery was not specifically assessed; however the above sample preparation and digestion protocol was thoroughly assessed for reproducibility during previous optimization and reproducibility studies.

Hydrophilic Interaction Liquid Chromatography (HILIC) peptide fractionation was performed on a Waters 1525 HPLC system equipped with a 4.6 × 25 mm TSK gel Amide-80 HR 5 μ m column (Tosoh Bioscience, South San Francisco, CA). CM (~900 μ g of protein) were loaded in 80% solvent B (98% acetonitrile, 0.1% TFA) and eluted with the following gradient: 80% B for 5 min followed by 80% B to 60% B in 40 min, 0% B in 5 min at 0.5 ml/min. Solvent A consisted of 98% HPLC grade water and 0.1% TFA collecting nine fractions.

Mass Spectrometry

Samples were analyzed by reverse-phase HPLC-ESI-MS/MS using an Eksigent Ultra Plus nano-LC 2D HPLC system (Dublin, CA) connected to a quadrupole time-of-flight TripleTOF 5600 (QqTOF) mass spectrometer (SCIEX). Typically, mass resolution for MS1 scans and corresponding precursor ions was ~35,000 (TripleTOF 5600), while resolution for MS/MS scans and resulting fragment ions (MRM-HR transitions) was ~15,000 ('high sensitivity' product ion scan mode) (Schilling et al., 2012). Briefly, after injection, peptide mixtures were transferred onto the analytical C18-nanocapillary HPLC column (C18 Acclaim PepMap100, 75 μ m I.D. × 15 cm, 3 μ m particle size, 100 Å pore size, Dionex, Sunnyvale, CA) and eluted at a flow rate of 300 nL/min using stepwise gradients from 5% to 80% solvent B with total runtimes, including mobile phase equilibration, of 90 min. Solvent mobile phase A was 2% acetonitrile/98% of 0.1% formic acid (v/v) in water, and mobile phase B was 98% acetonitrile/2% of 0.1% formic acid (v/v) in water. Data acquisition was performed in data dependent acquisition (DDA) mode on the TripleTOF 5600 to obtain MS/MS spectra for the 30 most abundant precursor ions (50 msec per MS/MS) following each survey MS1 scan (250 msec), yielding a total cycle time of 1.8 s as described (Kuhn et al., 2014; Schilling et al., 2012).

Bioinformatic Database Searches and SILAC Quantification

MS data were searched using the database search engine ProteinPilot (5) (SCIEX Beta 4.5, revision 1656) with the Paragon algorithm (4.5.0.0, 1654). The search parameters were set as follows: trypsin digestion, cysteine alkylation set to iodoacetamide, SILAC Quantification (Lys-6, Arg-6, no bias correction), and *Homo sapiens* as species. Trypsin specificity was assumed as C-terminal cleavage at lysine and arginine. Processing parameters were set to "Biological modification" and a thorough ID search effort was used. During the search, Protein Pilot performs an automatic mass recalibration of the datasets based on highly confident peptide spectra. Specifically, a first search iteration was done to select high confidence peptide identifications to recalibrate both the MS and MS/MS data, which is subsequently automatically re-searched. During the iterative steps of re-searching the data, the search parameters are less stringent, e.g., allowing for additional 'missed cleavages' (with typically not more than 2).

All data files were searched using the SwissProt 2014_01 release, released January 22, 2014), with a total of 40,464 human 'reviewed' protein sequences searched. A cut-off peptide confidence value of 99 was chosen, and a minimum of 2 identified peptides per protein was required. The Protein Pilot false discovery rate (FDR) analysis tool, the Proteomics System Performance Evaluation Pipeline (PSPEP algorithm) provided a global FDR of 1% and a local FDR at 1% in all cases. For protein quantification comparing proteins from 3 biological replicate experiments between CM from senescent (light) versus quiescent (heavy) cells a Protein Pilot significance threshold of < 0.05 was required. All database search results and details for peptide identifications and protein quantification are provided in Table S1.

Functional Analysis, Protein Ontology, Pathway Enrichment Analysis

For functional analysis and protein ontology analysis, 'The Database for Annotation, Visualization and Integrated Discovery' (DAVID v.6.7) was used (Huang et al., 2007). For pathway enrichment analysis, we used the Benjamini correction as adjustment to P values to determine statistical significance (Figures S1D–S1F).

Western Blotting

CM were collected and stored at -80°C , and cells remaining on the culture dish were counted for normalization. Protein concentrations were measured using Bradford reagents. Samples were incubated at 95°C for 10 min, loaded on 4%–15% gradient tris-glycine SDS-polyacrylamide gels (Invitrogen), separated by electrophoresis and transferred to PVDF membranes. Membranes were blocked in TBST/BSA for 1 h at room temperature, probed overnight at 4°C with primary antibodies in blocking buffer, washed in TBST and incubated with horseradish peroxidase-conjugated secondary antibodies for 1 h at room temperature. Signals were detected using Supersignal® West chemiluminescent substrate (Thermo Scientific). Densitometry was performed using ImageJ by the measurement of area under the curve method following background subtraction.

Platelet Activation Assay

Platelet-associated adenosine triphosphate (ATP) secretion was measured using the CHRONO-LUME reagent (Chrono-Log, Havertown, PA), according to the manufacturer's protocol with minor variations. Human platelet-rich plasma was incubated in CM from quiescent or IR-induced senescent HCA2 fibroblasts in the presence or absence of 5 μ g/mL collagen for 10 min prior to analysis. ATP was measured by adding CHRONO-LUME luciferin to platelet-rich plasma at a ratio of 1:9, after which luminescence was measured on a Perkin Elmer Victor X3 multilabel plate reader.

Isolation of Bone Marrow from Mouse Femurs

Femurs were isolated from both legs of each mouse and cut at top and bottom to uncover the bone marrow. The bones were flushed with cold PBS using a 25-gauge needle to collect the bone marrow into 1.6 mL Eppendorf tubes. Collected samples were centrifuged at $300 \times g$ for 5 minutes at 4°C . After centrifugation, the supernatant was removed, and the remaining pellet lysed in 1 mL TRIzol® reagent for total RNA isolation. RNA isolation was performed according to the manufacturer's instructions (Direct-zol RNA Miniprep Zymo Research cat# R2051).

Megakaryocyte Preparation

Megakaryocytes (MKs) were obtained and analyzed according to published protocols (Mathur et al., 2004; Schulze, 2012) with minor modifications. C57BL6 mice receiving different treatments were euthanized 8 days after treatment, and bone marrow cells were obtained from femurs and tibias by flushing. Single-cell suspensions were prepared by passage through 25-gauge needles, and collected by 5 min centrifugation at $200 \times g$. Cells were cultured in DMEM supplemented with 10% fetal bovine serum, 2 mM L-glutamine, penicillin/streptomycin, and 100 ng/ml murine thrombopoietin for 10 days. MK ploidy was measured by double staining with FITC conjugated anti-CD61 antibody and propidium iodide followed by flow cytometry on a BD FACS Aria II flow cytometer (Becton Dickinson). CD 61⁺ cells (regarded as MKs) were gated and analyzed for DNA content.

ELISAs

Platelet poor plasma from each treatment group was analyzed by ELISA according to the manufacturer's instructions. Thbs1 was from LifeSpan Biosciences, SerpinE1 (PAI-1) was from Abcam, uPA was from Innovative Research, and Timp1 was from R&D Systems. Plasma samples were diluted with Assay Buffers before analysis (Thbs1 = 1:5 dilution, PAI-1 = 1:5 dilution, uPA = 1:2 dilution, Timp1 = 1:2 dilution). Thrombopoietin (Tpo) was measured by ELISA (R&D Systems) on undiluted EDTA-treated plasma according to the manufacturer's instructions.

Quantitative PCR

RNA was extracted from cells using the ISOLATE II RNA mini kit (Bioline #BIO-52073), following the manufacturer's instructions. RNA was isolated from tissues via the Direct-zol RNA MiniPrep Kit (Zymo #11-331) according to the manufacturer's instructions. RNA (500 ng) was reverse-transcribed using the High Capacity cDNA Reverse Transcription Kit (Applied Biosystems #4368813). For each qPCR reaction, 10 ng of cDNA was used and quantified on a LightCycler 480 II (Roche) for 45 cycles. All signals were normalized to beta-actin.

FACS Analysis

Blood was collected from control and treated mice in TBS/Heparin (20U/ml), diluted in Tyrode's buffer, and centrifuged at $900 \times g$ for 5 min. The pellet was resuspended in Tyrode's buffer for further analysis. For measurement of platelet surface exposure of P-selectin or phosphatidylserine, we followed published protocols with minor modifications (Dachary-Prigent et al., 1993; Nieswandt et al., 2004). Briefly, washed blood was diluted 1:25 with Tyrode's buffer containing 1mM CaCl₂, left unstimulated or activated with 5 ug/ml type I collagen (Chrono-log) in the presence of a FITC-conjugated mouse anti-CD62P IgG (BD Biosciences) monoclonal antibody, Annex-V-APC (Biolegend, San Diego, CA). Within 30 min, samples were measured by flow cytometry on a FACS Aria II (BD Biosciences). No aggregation was observed in this time frame. For platelet counts, fluorescent beads (Thermo-Fisher #C36950) were added to volume equivalents of EDTA-treated blood diluted 1:1 in ice-cold PBS, which allowed normalization of platelet counts per unit volume.

Complete Blood Count (CBC), Activated Partial Thromboplastin Time (APTT), and Prothrombin Time (PT)

EDTA-treated blood (CBC) or citrated plasma (APTT/PT) was provided to a commercial research organization (Quality Veterinary Laboratory - Davis, CA). All assays were performed according to company protocols.

QUANTIFICATION AND STATISTICAL ANALYSIS

Error bars on all graphs show the standard error of the mean from at least three independent experiments for culture experiments, and at least 7 mice for animal experiments. The p values on single comparisons were performed by t test, while multiple comparisons were calculated using one-way ANOVA with Tukey's post-test. Numbers (n) for each experiment are indicated in figure legends, or individual sample plots are indicated. Significance is represented as * = $p < 0.05$, ** = $p < 0.01$, and *** = $p < 0.001$, with specific tests and significance markers indicated in each relevant figure.

DATA AND CODE AVAILABILITY

The mass spectrometric raw data associated with this manuscript can be downloaded from MassIVE at <http://MSV000079131@massive.ucsd.edu> (username: MSV000079131).